# jetMESSENGER® transfection reagent Short protocol - mRNA Transfection

### **DAY 0: Cell seeding**

→ Seed cells in V ml of serum containing medium according to the table below

#### Quantities per well, dish or flask

Culture vessel	Number of cells*	V = volume of growth medium for cell seeding
96-well	7 500 - 12 500	0.125 mL
24-well	12 000 - 50 000	0.5 mL
12-well	80 000 - 100 000	1 mL
6-well / 35 mm	150 000 - 200 000	2 mL
100 mm / flask 75 cm <sup>2</sup>	$2 \times 10^6 - 4 \times 10^6$	10 mL

<sup>\*</sup>For specific cell type or suspension cells, please refer to the complete protocol.

### **DAY 1: Transfection**

- → Perform transfection in the standard cell growth medium
- → Use jetMESSENGER® mRNA buffer only
- → Transfect cells at 60-80% confluency

Dilute **X** μg of mRNA in **W** μL of **mRNA buffer** Vortex 10 s and spin down

Add Y µL of jetMESSENGER® reagent (mRNA/jetMESSENGER® ratio 1:2)

Vortex 10 s, spin down and incubate 15 min at RT

Incubate 24 to 72 h

20 μL

Add transfection mix to the cells in serum containing medium

### Quantities per well, dish or flask

Culture vessel	W = volume of mRNA buffer	X = amount of mRNA added	Y = volume of jetMESSENGER® reagent
96-well	12.5 μL	<b>0.1</b> μg	0.25 μL
24-well	50 μL	0.5 μg	1 μL
12-well	100 μL	1 μg	2 μL
6-well / 35 mm	200 μL	<b>2</b> μg	4 μL

**10** μg

## **DAY 2-3: Measure gene expression**

See back page for optimization tips

100 mm / flask 75 cm<sup>2</sup>

Download complete protocol on <a href="http://www.polyplus-transfection.com/resources/product-literature/">http://www.polyplus-transfection.com/resources/product-literature/</a>

1000 µL



Contact us:

Phone: +33 (0)3 90 40 61 80

Email: support@polyplus-transfection.com Website: www.polyplus-transfection.com



Version C

# jetMESSENGER® transfection reagent Short protocol - Optimization Tips

## Protocol Optimization

- → Test different mRNA amounts between 0.5X and 2X.
- → Test different mRNA/jetMESSENGER® ratios, 1:2 to 1:3.
- ★ Check our online Cell Transfection Database for cell specific protocols at:

http://www.polyplus-transfection.com/resources/cell-transfection-database/



#### Quantities per well, dish or flask

Culture vessel	W volume of mRNA buffer	X amount of mRNA added	Y volume of jetMESSENGER® reagent
96-well	12.5 μL	$\textbf{0.1} \pm \textbf{0.05}~\mu\text{g}$	$0.25\pm0.05~\mu L$
24-well	50 μL	$\textbf{0.5} \pm \textbf{0.1}~\mu\text{g}$	$1\pm0.2~\mu$ L
12-well	100 μL	$1 \pm 0.2 \mu g$	$2 \pm 0.4~\mu$ L
6-well / 35 mm	200 μL	$2 \pm 0.5 \mu g$	$4\pm0.8~\mu$ L
100 mm / flask 75 cm <sup>2</sup>	1000 μL	$10 \pm 2.5  \mu g$	$20 \pm 4~\mu\text{L}$

## Tips to increase cell viability of sensitive cells

- ★ Wash cells 4 h after transfection.
- **★** Ensure that the mRNA is diluted in the mRNA buffer provided by Polyplus-transfection®.
- ♣ Analyze transfection at an earlier time point (e.g. at 24 h instead of 48 h).
- → Decrease the amount of mRNA added per well.
- → Decrease the volume of jetMESSENGER® reagent.
- → Use more stable chemically modified mRNA.
- → Check if the expressed protein may cause toxicity. If this is the case, reduce the amount of mRNA.

### Good mRNA Transfection Practices

- → Store appropriately jetMESSENGER® (5 ± 3°C) and the mRNA (- 80°C).
- → Ensure that the quality of your mRNA is optimal. Preferably use mRNA purchased from an oligo supplier, instead of homemade transcribed mRNA.
- → Use a common reporter gene-encoding mRNA as a positive control (ex: Luciferase or GFP).
- **★** Ensure the medium is permissive to the transfection.
- → The use of chemically modified mRNA (Pseudouridine, 5' Methylcytosine, 5-methoxyuridine, etc...) could improve the transfection efficiency.
- ★ Ensure that all reagents are RNAse-free.

**Note**: For more information regarding experimental conditions, please refer to the complete protocol available online at: http://www.polyplus-transfection.com/resources/product-literature/



